



A systematic review of the arsenic content of the normal human prostate gland

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Abstract

Background: The prostate gland is subject to various disorders. The etiology and pathogenesis of these diseases remain not well understood. Moreover, despite technological advancements, the differential diagnosis of prostate disorders has become progressively more complex and controversial. It was suggested that the arsenic (As) level in prostatic tissue plays an important role in prostatic carcinogenesis and its measurement may be useful as a cancer biomarker. These suggestions promoted more detailed studies of the As content in the prostatic tissue of healthy subjects.

Objective: The present study evaluated by systematic analysis the published data for as content analyzed in prostatic tissue of “normal” glands.

Methods: This evaluation reviewed 1927 studies, all of which were published in the years from 1921 to 2020 and were located by searching the databases Scopus, PubMed, MEDLINE, ELSEVIER-EMBASE, Cochrane Library, and the Web of Science. The articles were analyzed and “Median of Means” and “Range of Means” were used to examine heterogeneity of the measured As content in prostates of apparently healthy men. The objective analysis was performed on data from the 16 studies, which included 471 subjects.

Results: It was found that the range of means of prostatic as content reported in the literature for “normal” gland varies widely from 0.00039 mg/kg to <0.017 mg/kg with median of means 0.0031 mg/kg on a wet mass basis.

Conclusion: Because of small sample size and high data heterogeneity, we recommend other primary studies be performed.

Keywords: Arsenic, human prostate, normal prostatic tissue, biomarkers

Introduction

The prostate gland is subject to various disorders and of them chronic prostatitis, benign prostatic hyperplasia (BPH), and prostate cancer (PCa) are extremely common diseases of ageing men [1-3]. The etiology and pathogenesis of these diseases remain not well understood. A better understanding of the etiology and causative risk factors are essential for the primary prevention of these diseases.

In our previous studies the significant involvement of trace elements (TEs) in the function of the prostate was found [4-15]. It was also shown that levels of TEs in prostatic tissue can play a significant role in etiology of PCa [16-19]. Moreover, it was demonstrated that the changes of some TE levels, including arsenic (As), and TE content ratios in prostate tissue can be used as biomarkers [20-27].

The effects of TEs, including as, are related to their concentration. Recorded observations range from a deficiency state, through normal function as biologically essential components, to an imbalance, when excess of one element interferes with the function of another, to pharmacologically active concentrations, and finally to toxic and even life-threatening concentrations [28]. In this context, a significant dose-response relation was observed between low-level as exposure and cancers of the bladder, kidney, skin, and lung in both males and females, and cancers of the prostate and liver in males [29]. Recent available evidence in human populations and human cells

in vitro indicates that the prostate is a target for As carcinogenesis. A role for this common environmental contaminant in human PCa initiation and/or progression would be very important [30-36].

By now, an exceedingly scant literature exists on quantitative As content in tissue of “normal” and affected glands. The analyses reported are few in number, incomplete and difficult to interpret. Moreover, the findings of various studies indicate some discrepancies.

The present study addresses the significance of as levels in prostatic tissue as a biomarker of the gland's condition. Therefore, we systematically reviewed all the available relevant literature and performed a statistical analysis of as content in tissue of “normal” glands, which may provide valuable insight into the etiology and diagnosis of prostate disorders.

Materials and methods

Data sources and search strategy

Aiming at finding the most relevant articles for this review, a thorough comprehensive web search was conducted by consulting the Scopus, PubMed, MEDLINE, ELSEVIER-EMBASE, Cochrane Library, and the Web of Science databases, as well as from the personal archive of the author collected between May 1966 to September 2020, using the key words: prostatic trace elements, prostatic As content, prostatic tissue, and

their combinations. For example, the search terms for as content were: “As mass fraction”, “As content”, “As level”, “prostatic tissue as” and “As of prostatic tissue”. The language of the article was not restricted. The titles from the search results were evaluated closely and determined to be acceptable for potential inclusion criteria. Also, references from the selected articles were examined as further search tools. Relevant studies noted for the each selected article were also evaluated for inclusion.

Eligibility criteria

Inclusion criteria

Only papers with quantitative data of as prostatic content were accepted for further evaluation. Studies were included if the control groups were healthy human males with no history or evidence of urological or other Andrologia disease and as levels were measured in samples of prostatic tissue.

Exclusion criteria

Studies were excluded if they were case reports. Studies involving persons from as contaminated area and subjects that were as occupational exposed were also excluded.

Data extraction

A standard extraction of data was applied, and the following available variables were extracted from each paper: method of As determination, number and ages of healthy persons, sample preparation, mean and median of As levels, standard deviations of mean, and range of As levels. Abstracts and complete articles were reviewed independently, and if the results were different, the texts were checked once again until the differences were resolved.

Statistical analysis

Studies were combined based on means of as levels in prostatic tissue. The articles were analyzed and “Median of Means” and “Range of Means” were used to examine heterogeneity of as contents. The objective analysis was performed on data from the 16 studies, with 471 subjects.

Results

Information about as levels in prostatic tissue in different prostatic diseases is of obvious interest, not only to understand the etiology and pathogenesis of prostatic diseases more profoundly, but also for their diagnosis, particularly for PCa diagnosis and PCa risk prognosis [27]. Thus, it dictates a need for

reliable values of the As levels in the prostatic tissue of apparently healthy subjects, ranging from young adult males to elderly persons.

Possible publications relevant to the keywords were retrieved and screened. A total of 1927 publications were primarily obtained, of which 1911 irrelevant papers were excluded. Thus, 16 studies were ultimately selected according to eligibility criteria that investigated as levels in tissue of “normal” prostates (Table 1) and these 16 papers [9, 13, 14, 27, 37-48] comprised the material on which the review was based. A number of values for as mass fractions were not expressed on a wet mass basis by the authors of the cited references. However, we calculated these values using the medians of published data for water – 83% [49-52] and ash – 1% (on a wet mass basis) contents in “normal” prostates of adult men [51, 53-55].

Table 1 summarizes general data from the 16 studies. The retrieved studies involved 471 subjects. The ages of subjects were available for 10 studies and ranged from 0–87 years. Information about the analytical method and sample preparation used was available for 15 studies. Six studies determined as levels by destructive (require high temperature drying, acid digestion, fixation in ethanol/chloroform/formaldehyde, and paraffin/resin embedding of tissue samples) analytical methods (Table 1): one using atomic absorption spectrophotometry (AAS), two - inductively coupled plasma mass spectrometry (ICPMS), and three – radiochemical neutron activation analysis (RNAA). One study detected as level in intact prostatic tissue samples by nondestructive analytical method, such as instrumental neutron activation analysis (INAA). In eight studies a combination of destructive (ICPMS) and nondestructive (INAA) methods was used and results were summarized.

Discussion

The range of means of as mass fractions reported in the literature for “normal” prostatic tissue varies widely from 0.00039 mg/kg [27] to <2.9 mg/kg [37] with median of means <0.0031 mg/kg wet tissue (Table 1). The maximal value of mean as mass fraction reported [37] was 935 times higher the median of as mass fraction means and at least two orders of magnitude higher than all other published means. Thus, value <2.9 mg/kg [37] can be excluded. However, without this result range of means of as mass fractions for “normal” prostatic tissue remains very wide from 0.00039 mg/kg [27] to <0.017 mg/kg [38] with the same median of means <0.0031 mg/kg wet tissue and H_{max}/M_{min} ratio approximately 44 (Table 1).

Table 1: Reference data of as mass fractions (mg/kg wet tissue) in “normal” human prostatic tissue

Reference	Method	n	Age, years Range	Sample preparation	As	
					Mean±SD	Range
Zakutinsky <i>et al.</i> 1962 [37]	-	-	-	-	<2.9	-
Smith 1967 [38]	RNAA	10	Adult	D	0.0077	0.0017-0.0153
Liebscher <i>et al.</i> 1968 [39]	RNAA	10	Adult	D	0.0077±0.0037	0.0017-0.0153
Smith 1970 [40]	RNAA	10	Adult	D	0.0077±0.0037	0.0017-0.0153
Zaichick <i>et al.</i> 2012 [41]	ICP-MS	64	13-60	AD	≤0.0031	<0.0017-0.0275
Zaichick <i>et al.</i> 2013 [9]	2 methods	16	0-30	Intact, AD	≤0.012	-
Neslund-Dudas <i>et al.</i> 2014 [42]	ICP-MS	21 25	Adult, NS Adult, ES	F,P,AD,NB F,P,AD,NB	0.00092 0.00087	- -
Zaichick <i>et al.</i> 2014 [13]	2 methods	16	0-30	Intact, AD	≤0.017	-
Zaichick <i>et al.</i> 2014 [43]	2 methods	28	21-40	Intact, AD	≤0.0020	<0.0017-0.0034

		27	41-60	Intact, AD	≤ 0.0044	$< 0.0017-0.027$
		10	61-87	Intact, AD	0.0020	0.0017-0.0034
Zaichick <i>et al.</i> 2014 [14]	2 methods	16	0-30	Intact, AD	≤ 0.012	-
Zaichick <i>et al.</i> 2015 [44]	INAA	32	44-87	Intact	≤ 0.017	-
Zaichick 2015 [45]	2 methods	65	21-87	Intact, AD	≤ 0.0031	-
Zaichick <i>et al.</i> 2017 [46]	2 methods	37	41-87	Intact, AD	0.003	-
Zaichick 2017 [47]	2 methods	37	41-87	Intact, AD	≤ 0.0031	-
Singh <i>et al.</i> 2018 [27]	AAS	10	Adult	AD	0.00039 ± 0.00034	-
Zaichick <i>et al.</i> 2019 [48]	2 methods	37	41-87	Intact, AD	≤ 0.0031	-
Median of means					0.0031 or 0.0031 (without < 2.9)	
Range of means ($M_{\min} - M_{\max}$),					0.00039 - < 2.9 or $0.00039 < 0.017$ (without < 2.9)	
Ratio M_{\max}/M_{\min}					$< 2.9/0.00039 = < 7436$ or $< 0.017/0.00039 = < 43.6$ (without < 2.9)	
All references						16

M – arithmetic mean, SD – standard deviation of mean,

RNAA – radiochemical neutron activation analysis, ICPMS – inductively coupled plasma mass spectrometry; INAA – instrumental neutron activation analysis, AAS – atomic absorption spectrophotometry, 2 methods – INAA+ICPMS. NS – never-smokers, ES – ever-smokers, D – drying at high temperature, AD – acid digestion, F – fixed in ethanol/chloroform/formaldehyde, P – paraffin embedded, NB – needle biopsy.

This variability of reported mean values can be explained by a dependence of As content on many factors, including analytical method imperfections, differences in “normal” prostate definitions, possible non-homogeneous distribution of As levels throughout the prostate gland volume, age, ethnicity, diet, smoking, alcohol intake, consuming supplemental Zn and Se, and others. Not all these factors were strictly controlled in the cited studies. For example, in some studies the “normal” prostate means a gland of an apparently healthy man who had died suddenly, but without any morphological confirmation of “normality” of his prostatic tissue. In other studies the “normal” prostate means a non-cancerous prostate (but hyperplastic and inflamed glands were included) and even a visually “normal” prostatic tissue adjacent to a prostatic malignant tumor. In some studies whole glands were used for the investigation while in others the As content was measured in pieces of the prostate. However, the very short list of published data does not allowed us to estimate the effect of these factors on As content in “normal” prostate tissue.

In our opinion, the leading cause of inter-observer As content variability was insufficient quality control of results in published studies. Almost in all reported papers such destructive analytical methods as RNAA, AAS and ICPMS were used. These methods require acid digestion of the samples at a high temperature. There is evidence that use of this treatment causes some quantities of TEs to be lost [28, 56, 57]. Particularly, it concerns such volatile chemical element as As. On the other hand, the As content of chemicals used for acid digestion can contaminate the prostate samples. Thus, when using destructive analytical methods it is necessary to allow for the losses of TEs, for example when there is complete acid digestion of the sample. Then there are contaminations by TEs during sample decomposition, which require addition of some chemicals. In the case of a paraffin/epoxy embedded tissue samples As, particularly from prostatic fluid, may be lost during sample fixation in ethanol/chloroform/formaldehyde. It is possible to avoid these problems by using non-destructive methods, but up to now there are no analytical methods which allow to quantify As content in “normal” prostate without acid digestion of the samples at a high temperature. It is, therefore, reasonable to conclude that the quality control of results is very important factor for using the As content in prostatic tissue as biomarkers.

All natural chemical elements of the Periodic System, including As, present in all subjects of biosphere [28, 58, 59]. During the long evolutionary period intakes of As in organisms were more or less stable and organisms were adopted for such environmental conditions. Moreover, organisms, including human body, involved low doses of this element in their functions [60, 61]. As minerals have been known and used in relative small amounts since ancient times. As was frequently included in bronze, and used in medicine, cosmetics, and for murder. The situation began to change after the industrial revolution, particularly, over the last 100 years. The primary use of As is in industry, for example, in car batteries and ammunition, semiconductor electronic devices, optoelectronics, wood products as a wood preservative, pigments for plastics, ceramics and glasses. This metalloid is widely used in agriculture as pesticides, herbicides, and insecticides, as well as a feed additive in poultry and swine production. Its compounds are also used in medicine and military.

Thus, inorganic As is ubiquitously distributed in environment and food, water, and air everywhere contain this element. In addition to the abundant natural sources of As, there are a large number of industrial and agricultural sources of As to the soil (through atmospheric emissions originating from residues from coal, oil, and gas combustion, urban refuse, mine tailings, Au, Cu, and Pb smelter slag, waste, including pharmaceutical waste, smelting activities to phosphate fertilizers, and also form pesticides, herbicides, insecticides, and seaweed fertilizers application), water (through irrigation and industrial liquid waste, livestock dips, and wastewater sludge application), and air (through atmospheric industrial emissions) contamination. From the polluted environment As is subsequently introduced into the food chain [62]. However, the major source of human exposure to As on unpolluted territories is naturally contaminated drinking water from underground wells [28-30, 33, 34, 36].

Commercially, As is produced as As trioxide or as a pure element, however, these have not been produced in the United States since 1985. As trioxide is obtained as a byproduct from dusts and residues produced during the treatment of Au, Cu and Pb metal ores. China is the world’s leading producer of arsenic (25,000 tons in 2014) followed by Chile (10,000 tons), Morocco (8,000 tons), Russia (1,500 tons), Belgium (1,000 tons), Bolivia (52 tons), and Japan (45 tons). Since the use of As is linked to the rapidly developing modern technology, we can assume that over

the years, the need of industry in this metalloid has increased significantly and would continue to increase in the future. Published data showed an increase in As level for fluids and tissue of human body as the As intake increased [63-66]. Thus, we can conclude that the human body burden of As, including prostate tissue, has increased over the last 100 years due to an increase in global environmental As pollution [67]. It is likely that this tendency will continue.

As mentioned above, an ingestion of As by humans can cause a variety of disorders, such as skin lesions, problems with the respiratory and/or nervous systems, and different types of cancers, including PCa. Significant correlations between as exposure and the risk of PCa have been reported [29-36]. However, precise molecular mechanisms by which this metalloid causes healthy cells to transform to malignant states have yet to be fully defined. Kim *et al* [32], reported that inorganic As induces apoptosis, necrosis, and autophagy in the studied prostate cancer cell lines and suggest that this effect could be via a reactive oxygen species (ROS)-dependent mechanism. Tokar *et al* [68], showed that As can transform prostate epithelial stem cells into cancerous stem cells. Recent studies also reported the ability of As malignantly transforming human prostate epithelial cells via epigenetic alterations, such as miRNA dysregulation [69], silencing of mismatch repair gene MLH1 expression [70] while altering the expression of DNA methyl transferases such as DNMT1, DNMT3a, MeCP2, MBD1, and MBD4 [70, 71], and also genetic changes such as gene amplification leading to the overexpression of KRAS [72, 73].

Thus, according our study for unpolluted areas there are no information could explain the variability of published means for “normal” prostatic As levels from 0.00039 mg/kg to <0.017 mg/kg in wet tissue. Moreover, prostate tissue as contents showed large variations among individuals, but sources of the variation remain unknown. It is, therefore, reasonable to assume from data of our study that inaccuracy of analytical technologies employed caused so great variability of published means for prostatic as levels. This conclusion was supported the fact that the Certified Reference Materials for quality control of results were used only in a very few reported studies.

There are some limitations in our study, which need to be taken into consideration when interpreting the results of this review. The sample size of each study was sometimes relatively small (from 10 to 65), and a total of 471 “normal” prostates were investigated from all 16 studies. As such, it is hard to draw definite conclusions about the reference value of the As content in “normal” prostate as well as about the clinical value of the As levels in “normal” prostates as a biomarker.

Conclusions

The present study is a comprehensive study regarding the determination of as content in “normal” human prostates. With this knowledge as levels may then be considered as a biomarker for the recognition of prostate disorders. The study has demonstrated that levels of as in “normal” prostates depends on many unknown factors. Because of the uncertainties we have outlined, we recommend other primary studies be performed.

Competing Interests

Author has declared that no competing interests exist. The author

alone is answerable for the substance and composing of the paper. The author did not receive any funds from any source.

References

- Nickel JC. Prostatitis. *Can Urol Assoc J.* 2011; 5:306-315.
- Lim KB. Epidemiology of clinical benign prostatic hyperplasia. *Asian J Urol.* 2017; 4:148-151
- Rawla P. Epidemiology of Prostate Cancer. *World J Oncol.* 2019; 10(2):63-89.
- Avisyn AP, Dunchik VN, Zhavoronkov AA, Zaichick VE, Sviridova TV. Histological structure of the prostate and content of zinc in it during various age period. *Archiv Anatomy, Gistology, and Ebriology (Leningrad).* 1981; 81(11):76-83.
- Zaichick V. INAA and EDXRF applications in the age dynamics assessment of Zn content and distribution in the normal human prostate. *J Radioanal Nucl Chem.* 2004; 262:229-234
- Zaichick V, Zaichick S. The effect of age on Br, Ca, Cl, K, Mg, Mn, and Na mass fraction in pediatric and young adult prostate glands investigated by neutron activation analysis. *Appl Radiat Isot.* 2013; 82:145-151.
- Zaichick V, Zaichick S. INAA application in the assessment of Ag, Co, Cr, Fe, Hg, Rb, Sb, Sc, Se, and Zn mass fraction in pediatric and young adult prostate glands. *J Radioanal Nucl Chem.* 2013; 298:1559-1566.
- Zaichick V, Zaichick S. NAA-SLR and ICP-AES application in the assessment of mass fraction of 19 chemical elements in pediatric and young adult prostate glands. *Biol Trace Elem Res.* 2013; 156:357-366.
- Zaichick V, Zaichick S. Use of neutron activation analysis and inductively coupled plasma mass spectrometry for the determination of trace elements in pediatric and young adult prostate. *Am J Analyt Chem.* 2013; 4:696-706.
- Zaichick V, Zaichick S. Relations of bromine, iron, rubidium, strontium, and zinc content to morphometric parameters in pediatric and no hyperplastic young adult prostate glands. *Biol Trace Elem Res.* 2014; 157:195-204.
- Zaichick V, Zaichick S. Relations of the neutron activation analysis data to morphometric parameters in pediatric and no hyperplastic young adult prostate glands. *Advances in Biomedical Science and Engineering.* 2014; 1:26-42.
- Zaichick V, Zaichick S. Relations of the Al, B, Ba, Br, Ca, Cl, Cu, Fe, K, Li, Mg, Mn, Na, P, S, Si, Sr, and Zn mass fractions to morphometric parameters in pediatric and nonhyperplastic young adult prostate glands. *Bio Metals* 2014; 27:333-348.
- Zaichick V, Zaichick S. The distribution of 54 trace elements including zinc in pediatric and nonhyperplastic young adult prostate gland tissues. *Journal of Clinical and Laboratory Investigation Updates.* 2014; 2(1):1-15.
- Zaichick V, Zaichick S. Androgen-dependent chemical elements of prostate gland. *Androl Gynecol: Curr Res.* 2014; 2:2.
- Zaichick V, Zaichick S. Differences and relationships between morphometric parameters and zinc content in nonhyperplastic and hyperplastic prostate glands. *Br J Med & Med Res.* 2015; 8:692-706.
- Schwartz MK. Role of trace elements in cancer. *Cancer Res.*

- 1975; 35:3481-3487.
17. Zaichick V, Zaichick S. Role of zinc in prostate cancerogenesis. In: Mengen und Spurenelemente. 19. Arbeitstagung. Friedrich-Schiller-Universität, Jena, 1999, 104-115.
 18. Zaichick V, Zaichick S, Wynchank S. Intracellular zinc excess as one of the main factors in the etiology of prostate cancer. *J Anal Oncol.* 2016; 5:124-131.
 19. Zaichick V, Zaichick S, Rossmann M. Intracellular calcium excess as one of the main factors in the etiology of prostate cancer. *AIMS Mol Sci.* 2016; 3:635-647.
 20. Dunchik V, Zherbin E, Zaichick V, Leonov A, Sviridova T. Method for differential diagnostics of prostate malignant and benign tumours. Russian patent (Author's Certificate No 764660, priority of invention 27.10.1977). Discoveries, Inventions, Commercial Models, Trade Marks. 1980; 35:13.
 21. Zaichick V, Sviridova T, Zaichick S. Zinc in the human prostate gland: normal, hyperplastic and cancerous. *Int Urol Nephrol.* 1997; 29:565-574.
 22. Zaichick V, Sviridova T, Zaichick S. Zinc in human prostate gland: normal, hyperplastic and cancerous. *J Radioanal Nucl Chem.* 1997; 217:157-161
 23. Zaichick S, Zaichick V. Trace elements of normal, benign hypertrophic and cancerous tissues of the human prostate gland investigated by neutron activation analysis. *J Appl Radiat Isot.* 2012; 70:81-87.
 24. Zaichick V, Zaichick S. Ratios of selected chemical element contents in prostatic tissue as markers of malignancy. *Hematol Med Oncol.* 2016; 1(2):1-8.
 25. Zaichick V, Zaichick S. Trace element levels in prostate gland as carcinoma's markers. *J Cancer Ther.* 2017; 8:131-145
 26. Zaichick V, Zaichick S. Ratios of Zn/trace element contents in prostate gland as carcinoma's markers. *Cancer Rep Rev.* 2017; 1(1):1-7.
 27. Singh CK, Sinha P, Anshu AK. High accumulation of arsenic in prostate cancer patients in Gangetic zone of Bihar. *IOSR Journal of Biotechnology and Biochemistry.* 2018; 4(2):1-4.
 28. Zaichick V. Medical elementology as a new scientific discipline. *J Radioanal Nucl Chem.* 2006; 269:303-309.
 29. Wu MM, Kuo TL, Hwang YH, Chen CJ. Dose-response relation between arsenic concentration in well water and mortality from cancers and vascular diseases. *Am J Epidemiol.* 1989; 130(6):1123-1132.
 30. Lewis DR, Southwick JW, Ouellet-Hellstrom R, Rench J, Calderon RL. Drinking water arsenic in Utah: A cohort mortality study. *Environ Health Perspect.* 1999; 107(5):359-365.
 31. Benbrahim-Tallaa L, Waalkes MP. Inorganic Arsenic and Human Prostate Cancer. *Environ Health Perspect.* 2008; 116(2):158-164.
 32. Kim Y, Jeong IG, Ypu D, Hoon S, Sduh SN, Jang S-W, *et al.* Sodium meta-arsenite induces reactive oxygen species-dependent apoptosis, necrosis, and autophagy in both androgen-sensitive and androgen-insensitive prostate cancer cells. *Anticancer Drugs.* 2014; 25(1):53-62.
 33. Bulka CM, Jones RM, Turyk ME, Stayner LT, Argos M. Arsenic in drinking water and prostate cancer in Illinois counties: An ecologic study. *Environ Res.* 2016; 148:450-456.
 34. Roh T, Lynch CF, Weyer P, Wang K, Kelly KM, Ludewig G *et al.* Low-level arsenic exposure from drinking water is associated with prostate cancer in Iowa. *Environ Res.* 2017; 159:338-343.
 35. Lundqvist J, Helmersson E, Oskarsson A. Hormetic dose response of NaAsO₂ on cell proliferation of prostate cells *in vitro*: Implications for prostate cancer initiation and therapy. *Dose Response.* 2019; 17(2):1559325819843374
 36. Ahn J, Boroje II, Ferdosi H, Kramer ZJ, Lamm SH. Prostate cancer incidence in U.S. counties and low levels of arsenic in drinking water. *Int J Environ Res Public Health.* 2020; 17(3):960
 37. Zakutinsky DI, Parfyenov YuD, Selivanova LN. Data book on the radioactive isotopes toxicology. State Publishing House of Medical Literature, Moscow, 1962.
 38. Smith H. The distribution of antimony, arsenic, copper, and zinc in human tissue. *J Forensic Science.* 1967; 7:97-102.
 39. Liebscher K, Smith H. Essential and nonessential trace elements. A method of determining whether an element is essential or nonessential in human tissue. *Arch. Environ. Health.* 1968; 17:882-891.
 40. Smith H. Determination of trace elements in biological material by neutron activation analysis. In: Trace Element Metabolism in Animals. Proceedings WAAP/IBP International Symposium (July 1969, Aberdeen, Scotland). E. & S. Livingstone, Edinburg-London, 1970, 512-521.
 41. Zaichick S, Zaichick V, Nosenko S, Moskvina I. Mass fractions of 52 trace elements and zinc trace element content ratios in intact human prostates investigated by inductively coupled plasma mass spectrometry. *Biol Trace Elem Res.* 2012; 149:171-183.
 42. Neslund-Dudas C, Kandedgedara A, Kryvenko ON, Gupta N, Rogers C, Rybicki BA, Ping Dou Q, Mitra B. Prostate tissue metal levels and prostate cancer recurrence in smokers. *Biol Trace Elem Res.* 2014; 157:107-112.
 43. Zaichick V, Zaichick S. Use of INAA and ICP-MS for the assessment of trace element mass fractions in adult and geriatric prostate. *J Radioanal Nucl Chem.* 2014; 301:383-397.
 44. Zaichick V, Zaichick S, Davydov G. Differences between chemical element contents in hyperplastic and nonhyperplastic prostate glands investigated by neutron activation analysis. *Biol Trace Elem Res.* 2015; 164:25-35.
 45. Zaichick V. The Variation with Age of 67 Macro- and Microelement Contents in Nonhyperplastic Prostate Glands of Adult and Elderly Males Investigated by Nuclear Analytical and Related Methods. *Biol Trace Elem Res.* 2015; 168:44-60.
 46. Zaichick V, Zaichick S. Chemical Element Contents in Normal and Benign Hyperplastic Prostate. *Ann Mens Health Wellness.* 2017; 1(2):1006.
 47. Zaichick V. Differences between 66 Chemical Element Contents in Normal and Cancerous Prostate. *Journal of Analytical Oncology.* 2017; 6:37-56.
 48. Zaichick V, Zaichick S. Comparison of 66 chemical element contents in normal and benign hyperplastic prostate. *Asian J*

- Urol. 2019; 6:275-289.
49. Isaacs JT. Prostatic structure and function in relation to the etiology of prostatic cancer. *Prostate*. 1983; 4(4):351-366.
 50. Leissner KM, Fielkegard B, Tisell LE. Concentration and content of zinc in human prostate. *Invest Urol*. 1980; 18:32-35.
 51. Woodard HQ, White DR. The composition of body tissues. *Br J Radiol*. 1986; 59:1209-1218.
 52. Arnold WN, Thrasher JB. Selenium concentration in the prostate. *Biol Trace Elem Res*. 2003; 91(3):277-280.
 53. Tipton IH, Cook MJ. Trace elements in human tissue. Part II. Adult subjects from the United States. *Health Phys*. 1963; 9(2):103-145.
 54. Schroeder HA, Nason AP, Tipton IH, Balassa JJ. Essential trace metals in man: Zinc. Relation to environmental cadmium. *J Chron Dis*. 1967; 20:179-210.
 55. Saltzman BE, Gross SB, Yeager DW, Meiners BG, Gartside PS. Total body burdens and tissue concentrations of lead, cadmium, copper, zinc, and ash in 55 human cadavers. *Environ Res*. 1990; 52:126-145.
 56. Zaichick V. Sampling, sample storage and preparation of biomaterials for INAA in clinical medicine, occupational and environmental health. In: *Harmonization of Health-Related Environmental Measurements Using Nuclear and Isotopic Techniques*. IAEA, Vienna, 1997, 123-133.
 57. Zaichick V. Losses of chemical elements in biological samples under the dry ashing process. *Trace Elements in Medicine (Moscow)*. 2004; 5(3):17-22.
 58. Vernadsky VI. *Living Matter*, Nauka, Moscow, 1978.
 59. Zaichick V, Ermidou-Pollet S, Pollet S. Medical elementology: a new scientific discipline. *Trace Elements and Electrolytes*. 2007; 24(2):69-74.
 60. Anke M, Müller M, Hoppe C. Recent progress in exploring the essentiality of the ultratrace element aluminum to the nutrition of animals and man. *Biomed Res Trace Elem*. 2005; 16(3):183-187.
 61. Uthus EO. Arsenic essentiality: A role affecting methionine metabolism: *The Journal of Trace Elements in Experimental Medicine*. 2003; 16(4):345-355.
 62. Punshon T, Jackson BP, Meharg AA, Warczack T, Scheckel K, Guerinot ML *et al*. Understanding arsenic dynamics in agronomic systems to predict and prevent uptake by crop plants. *Sci Total Environ*. 2017; 581-582:209-220.
 63. Valentine JI, Kang HK, Spivey G. Arsenic levels in human blood, urine, and hair in response to exposure via drinking water. *Environ. Res*. 1979; 20(1):24-32.
 64. Adair BM, Moore T, Conklin SD, Creed JT, Wolf DC, David J Thomas DJ *et al*. Tissue distribution and urinary excretion of dimethylated arsenic and its metabolites in dimethylarsinic acid- or arsenate-treated rats. *Toxicol Appl Pharmacol*. 2007; 222(2):235-242
 65. Drobná Z, Walton FS, Paul DS, Xing W, Thomas DJ, Stýblo M *et al*. Metabolism of arsenic in human liver: the role of membrane transporters. *Arch Toxicol*. 2010; 84(1):3-16
 66. Hata A, Kurosawa H, Endo Y, Yamanaka K, Fujitani N, Endo G. A biological indicator of inorganic arsenic exposure using the sum of urinary inorganic arsenic and monomethylarsonic acid concentrations. *J Occup Health*. 2016; 58(2):196-200.
 67. Chung JY, Yu SD, Hong YS. Environmental source of arsenic exposure. *J Prev Med Public Health*. 2014; 47(5):253-257.
 68. Tokar EJ, Diwan BA, Waalkes MP. Arsenic exposure transforms human epithelial stem/progenitor cells into a cancer stem-like phenotype. *Environ Health Perspect*. 2010; 118:108-115.
 69. Cardoso APF, Al-Eryani L, States JC. Arsenic-induced carcinogenesis: the impact of miRNA dysregulation. *Toxicol. Sci*. 2018:165:284-290.
 70. Treas J, Tyagi T, Singh KP. Chronic exposure to arsenic, estrogen, and their combination causes increased growth and transformation in human prostate epithelial cells potentially by hypermethylation-mediated silencing of MLH1. *Prostate*. 2013; 73:1660-1672.
 71. Bustaffa E, Stocco A, Bianchi F, Migliore L. Genotoxic and epigenetic mechanisms in arsenic carcinogenicity. *Arch Toxicol*. 2014; 88:1043-1067.
 72. Merrick BA, Phadke DP, Bostrom MA, Shah RR, Wright GM, Wang X *et al*. Arsenite malignantly transforms human prostate epithelial cells *in vitro* by gene amplification of mutated KRAS. *PLoS One*. 2019; 14:e0215504.
 73. Lim JT, Tan YQ, Valeri L, Lee J, Geok PP, Chia SE *et al*. Association between serum heavy metals and prostate cancer risk—A multiple metal analysis. *Environ Int*. 2019; 132:105109.